

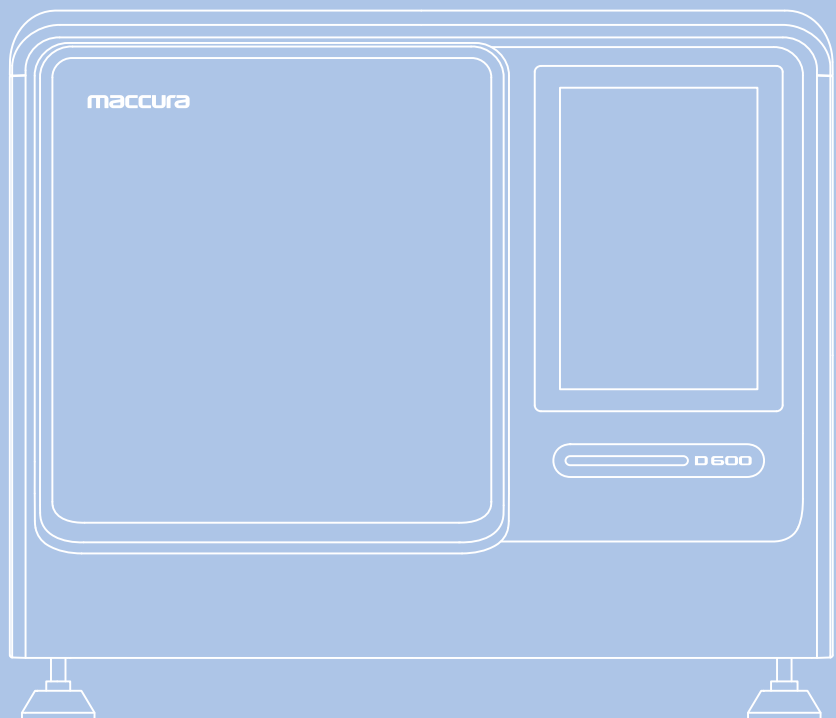
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современная лаборатория

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maccura

D600

DIGITAL PCR SYSTEM



About Digital PCR

Digital PCR(dPCR) technology is an absolute quantitative nucleic acid detection method, without the need for standard curve to accurately quantify nucleic acid, greatly ensuring the accuracy and precision of detection, is the third generation of quantitative nucleic acid detection technology after real-time fluorescence quantitative PCR technology. It has important application value in liquid biopsy, tumor concomitant diagnosis, noninvasive prenatal screening, pathogen load monitoring and so on.



Absolute
quantification

No reference required
Independent of the
standard curve



Accurate and
sensitive

High signal-to-noise ratio
Single copy can be tested



Reliable
retest

Good repeatability
Ensure the consistency of
test results



High efficiency
and stability

Tolerance to inhibitors
Applicable to complex
samples

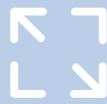
Original Vibration Micro(OsciDrop®) Core Technology

D600 automatic digital PCR system, with self-developed core technology of vibrating microdroplet, has many breakthrough advantages.



Fully automatic

The whole process of sampling, droplet generation, amplification and testing is realized.



High throughput

96 samples in three hours
Suitable for different
throughput requirements



Low cost

Chip-free consumables
Cost is reduced to qPCR
level



Multiple channels

Up to 6 channel fluorescence
channels
The product can be recycled
separately



High precision

In situ amplification and testing
Improve accuracy and precision



Customization

Number of nanodroplets
per sample
It can be customized
according to testing
requirements

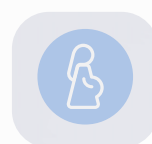
Applications



Pathogen



Oncology



NIPT



Environment



Minimal Residual
Detection



Copy Number
Variation



Gene Editing



Immunotherapy



Food Safety



Epigenetics



Gene
Expression



Single Cell
Research

Parameters

| | |
|------------|------------------------------------|
| Instrument | D 600 Automatic Digital PCR System |
|------------|------------------------------------|

| | |
|---------------------------|----------------------------------|
| Reaction wells per sample | 5,000-20,000 nanodroplets/sample |
|---------------------------|----------------------------------|

| | |
|------------------|------|
| Nanodroplet size | 1 nL |
|------------------|------|

| | |
|-------------------|--------------|
| Sample throughput | 1-96 samples |
|-------------------|--------------|

| | |
|-----------------|-------------------------------|
| Optical Channel | Up to 6 fluorescence channels |
|-----------------|-------------------------------|

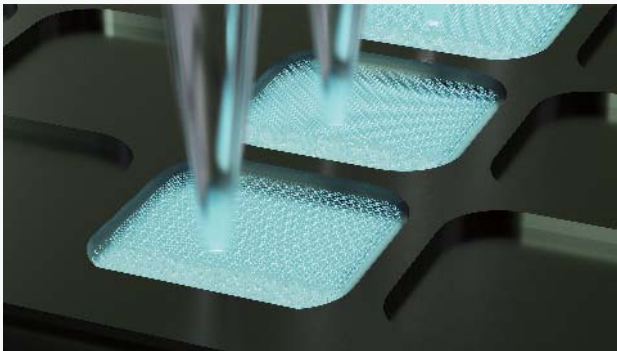
| | |
|-------------|-------------|
| Sensitivity | Single copy |
|-------------|-------------|

| | |
|---------------|--------|
| Dynamic range | 5 logs |
|---------------|--------|

| | |
|-----------|------|
| Precision | ±10% |
|-----------|------|

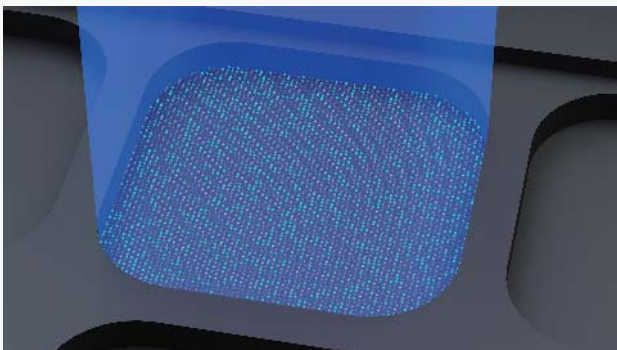
| | |
|---------------------|-----|
| Sample illumination | LED |
|---------------------|-----|

| | |
|------------------|------|
| Sample detection | CMOS |
|------------------|------|



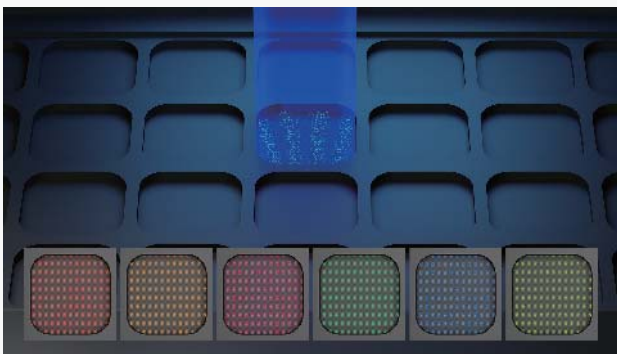
Fast & uniformity

Microdroplet array printing adopts the original vibration micro(OsciDrop®) core technology to rapidly and stably generate uniform nano-scale microdroplet.



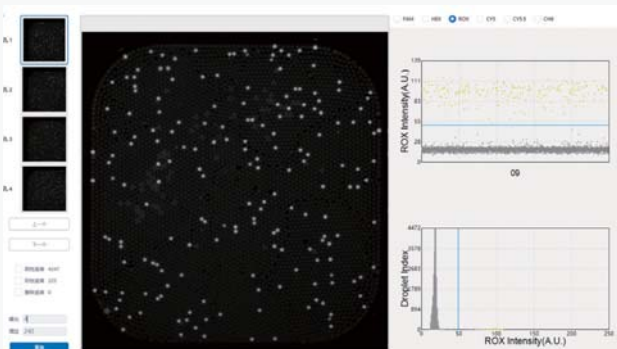
Stable amplification

The micro nanodroplets are flattened into a single-layer array in the well plate for in-situ amplification and fluorescence reading detection. The operation is convenient and the nanodroplets are effectively used.



Multiple fluorescence

Nanodroplet array reading has high-performance LED excitation light source and orthogonal fluorescence imaging detection system, supports more fluorescence detection channels, and can develop super multiplex digital PCR detection system.



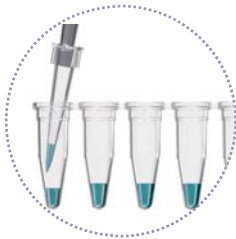
Exact algorithm

Using advanced planapochromatic fluorescence imaging correction technology and deep learning nanodroplet array recognition algorithm, the nanodroplet recognition accuracy is high and the signal-to-noise ratio is good.

* The pictures are for visual effects, for illustration and expression only, not for actual reference.

One-stop detection process

Step 1



Configure PCR reaction system

Reaction system: 10 ~ 25 μ L

Manual configuration of the reaction system

Step 2



automatic detection and analysis

Nanodroplet generation

Put the reaction system and consumables into the instrument for nanodroplet generation. Each experiment can generate up to 96 nanodroplets of samples; Each print and sample can generate up to 20,000 nanodroplets.

PCR amplification reaction

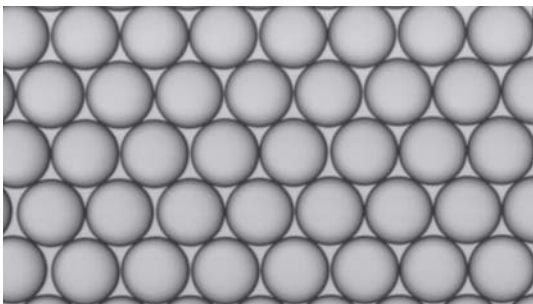
After the nanodroplets are generated, the instrument seals the amplification area of the orifice plate to complete the PCR reaction.

Result analysis

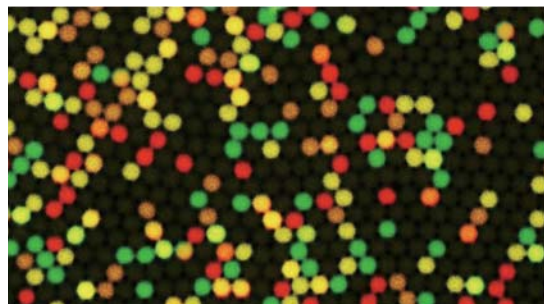
Fluorescence photo-detection is performed on each reaction well; the quantitative value of the sample is calculated according to the number of negative and positive nanodroplets, which is according to poisson distribution.

Product performance

The nanodroplet size is accurate and uniform nanodroplet amplification is rapid and stable



OsciDrop® nanodroplet
generation is uniform and stable



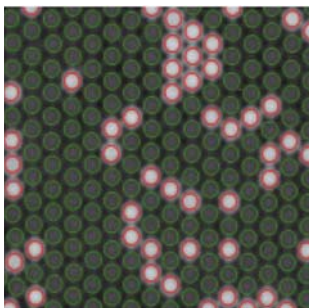
OsciDrop@4 fluorescence channels test
results

OsciDrop® provides a new platform for the wide range of applications of digital PCR adopting Maccura chip-free vibration micro(OsciDrop®) core technology, combined with advanced micro-nano sample needle, new fluorescent-free drop-generating oil, and high-performance digital PCR premixed solution.

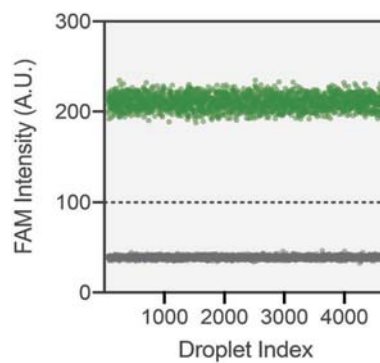
Original vibration micro(OsciDrop®) core technology, precisely set the volume of each microdroplet, volume CV(coefficient of variation) value is less than 5%, meets the requirements of digital PCR accurate quantification.

High performance flat plate amplification platform, combined with high thermal conductivity microdroplet array porous plate, improve the uniformity of large area nanodroplet array PCR rapid amplification.

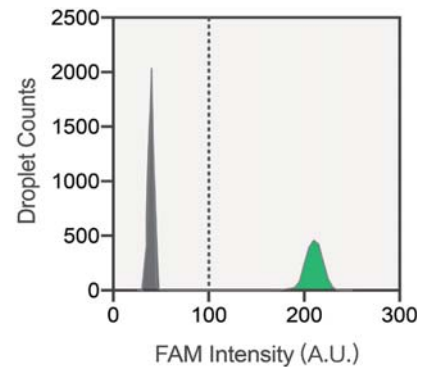
Ultra-sensitive multi-channel fluorescence imaging
Nanodroplet signal interpretation is accurate and fast



AI based nanodroplet interpretation



Scatter diagram of nanodroplet signal



Histogram of nanodroplet signal

Super Hi-Vision orthogonal multi-channel fluorescence imaging technology, can achieve up to 6 color channel fluorescence imaging, high detection sensitivity, effectively avoid fluorescence channel crosstalk.

The absolute quantification of nucleic acid in the experimental samples shows that there was no raining phenomenon in the positive and negative droplets, and the signal-to-noise ratio was high. D600 automatic digital PCR system supports analysis software enables users to view the original fluorescence map of microdroplet array, software interpretation results and quantitative statistical results.

*The red circle is the nanodroplet judged positive by the software, and the green circle is the negative nanodroplet.

Detection Kit

Instrument D 600 Automatic Digital PCR System

Consumable Universal dPCR Consumables

Universal Reagent 2× Universal dPCR MasterMix for DNA
2× One Step RT-dPCR MasterMix for RNA

Customized Tests Menu

HIV-1 Digital PCR Kit
2019-nCoV Digital PCR Kit
Influenza A Virus Digital PCR Kit
Influenza B Virus Digital PCR Kit
Rhinovirus Digital PCR Kit
Adenovirus Digital PCR Kit
Mycoplasma Pneumoniae Digital PCR Kit
Mycobacterium Tuberculosis Digital PCR Kit
Norovirus Digital PCR Kit
Enterovirus 71 Digital PCR Kit
Enterovirus Universal Digital PCR Kit
Coxsackie Virus A 16 Digital PCR Kit
Monkeypox Virus Digital PCR kit
Human EGFR T790M Digital PCR Kit
Human PIK3CA E542K Digital PCR Kit

Extraction N32 Automatic Extraction System
Nucleic acid extraction or purification kit

* For research use only. Digital PCR kits provide various customized services

Customized service

In addition to supporting kits, it can provide customization options, including infection diseases, tumor markers, genetic and other related diseases, a total of more than 400, please refer to the form on the next page for details.

400⁺ Continuously add

Infection related

Respiratory pathogens
Hand, foot, mouth and enterovirus pathogens
Hepatitis virus
Reproductive health
Parasite

Tumor related

Mutation detection
Copy number variation
Fusion gene detection
Methylation detection

Genetically related

Single gene disease

Animal related

Animal disease
Zoonotic disease
Pets

D600

Automatic Digital PCR System Customized Reagents

| | | |
|-------------------------|--|---|
| Flu A | Nucleic acid typing of PIV | <i>Nocardia asteroides</i> |
| Flu A H1N1 Virus (2009) | PIV- I | <i>Nocardia brasiliensis</i> |
| Flu A H3N2 subtype | PIV- II | <i>Nocardia transvalensis</i> |
| Flu B | PIV- III | <i>Nocardia carnea</i> |
| Flu B Victoria | PIV- IV | <i>Cryptococcus neoformans</i> |
| Flu B Yamagata | HCoV | <i>Haemophilus flu B</i> |
| Flu N | HCoV typing | <i>Streptococcus suis</i> |
| Flu N1 | HCoV 229E | <i>Chryseobacterium</i> |
| Flu N2 | HCoV OC43 | <i>meningosepticum</i> |
| Flu N3 | HCoV NL63 | <i>Neisseria meningitidis</i> |
| Flu N4 | HCoV HKU1 | <i>Neisseria meningitidis serogroup</i> |
| Flu N5 | MERS-CoV | <i>W135</i> |
| Flu N6 | SARS-CoV | <i>Neisseria meningitidis serogroup Y</i> |
| Flu N7 | 2019-nCoV (ORF1ab/E) | <i>Neisseria meningitidis serogroup X</i> |
| Flu N8 | 2019-nCoV (ORF1ab/N) | <i>Neisseria meningitidis serogroup C</i> |
| Flu N9 | 2019-nCoV (ORF1ab/E/N) | <i>Neisseria meningitidis serogroup B</i> |
| Flu H1 | 2019-nCoV and N501Y mutation | <i>Neisseria meningitidis serogroup A</i> |
| Flu H2 | Mutations in the S gene HV69-70del, E484K and K417N of 2019-nCoV | <i>Group A+C Neisseria meningitidis</i> |
| Flu H3 | Mutations in the S gene L452R, E484Q, P681R of 2019-nCoV | <i>Acinetobacter baumannii</i> |
| Flu H4 | ADV | <i>Enterovirus D68v</i> |
| Flu H5 | ADV- III | <i>Moraxella cataracea</i> |
| Flu H6 | ADV- VII | <i>KI polyoma virus</i> |
| Flu H7 | HRV | <i>Aeromonas</i> |
| Flu H8 | HMPV | <i>Corynebacterium diphtheria</i> |
| Flu H9 | HBoV | <i>Legionella micdadei</i> |
| Flu H10 | Mumps virus | <i>Pseudomonas aeruginosa</i> |
| Flu H11 | WU polyoma virus | <i>Group A Streptococcus</i> |
| Flu H12 | <i>Streptococcus pneumoniae</i> | <i>Group B streptococcus</i> |
| Flu H13 | <i>Haemophilus influenzae</i> | <i>Aeromonas hydrophila</i> |
| Flu H14 | <i>Staphylococcus aureus</i> | <i>Yersinia pseudotuberculosis</i> |
| Flu H15 | <i>Group A B hemolytic streptococcus</i> | <i>Drug-resistant Pseudomonas aeruginosa</i> |
| Flu H16 | <i>Ebola Virus</i> | <i>aeruginosa</i> |
| AI virus | <i>Klebsiella pneumoniae</i> | <i>MRSA</i> |
| AI virus H5N1 | <i>Chlamydia pneumoniae</i> | <i>Extended spectrum beta-lactamases bacteria</i> |
| AI virus H5N2 | <i>Mycoplasma pneumoniae</i> | <i>Stenotrophomonas maltophilia</i> |
| AI virus H5N6 | <i>Legionella bacteria</i> | <i>Mycobacterium leprae</i> |
| AI virus H5N8 | <i>Legionella pneumophila</i> | <i>Mycobacterium abscessus</i> |
| AI virus H7N2 | <i>Pseudomonas aeruginosa</i> | <i>Mycobacterium intracellulare</i> |
| AI virus H7N4 | <i>Mycobacterium tuberculosis</i> | <i>Mycobacterium avium</i> |
| AI virus H7N9 | <i>Nocardia asteroides</i> | <i>Mycobacterium gordonae</i> |
| AI virus H9N2 | <i>Bordetella pertussis</i> | <i>Mycobacterium abscessus</i> |
| AI virus H10N8 | <i>Bacillus parapertussis</i> | <i>Mycobacterium kansasii</i> |
| Flu C | <i>Corynebacterium diphtheriae</i> | <i>Rhodococcus equi</i> |
| Flu D | <i>Pneumocystis jirovecii</i> | <i>Clostridium tetani</i> |
| RSV | | |
| RSV A | | |
| RSV B | | |
| PIV | | |

| | | |
|----------------------------------|----------------------------------|-------------------------|
| Enterovirus universal | SED | Bacillus cereus |
| Enterovirus 71 | SEE | Clostridium perfringens |
| Coxsackie virus A16 | ECHO 11 | Cryptosporidium |
| Coxsackie virus A6 | ECHO 30 | Entamoeba histolytica |
| Coxsackie virus A10 | Botulinum toxin | Giardia |
| Enterovirus 68 | Botulinum toxin A | Human parechovirus |
| Enterovirus 70 | Botulinum toxin B | Listeria monocytogenes |
| Coxsackie virus A2 | Botulinum toxin E | Campylobacter coli |
| Coxsackie virus A4 | Botulinum toxin F | Salmonella typhi |
| Coxsackie virus A5 | Yersinia enterocolitica | paratyphosus bacillus |
| Coxsackie virus A24 | Yersinia enterocolitis virulence | |
| Coxsackie virus B1 | gene | |
| Coxsackie virus B2 | EAEC | |
| Coxsackie virus B3 | EPEC | |
| Coxsackie virus B4 | ETEC | |
| Coxsackie virus B5 | EHEC | |
| Coxsackie virus B6 | EIEC | |
| Norovirus * | E.coli universal | |
| Norovirus G I | EHEC O157 | |
| Norovirus G II | EHEC O104 | |
| Norovirus IV | Hemolysin gene | |
| Poliovirus 1 | E. coli stx1 gene | |
| Poliovirus 2 | E. coli stx2 gene | |
| Poliovirus 3 | Astrovirus | |
| RV A | Sapovirus | |
| RV B | Enteric Adenovirus | |
| RV C | Parechovirus | |
| Vibrio cholera | Picobirnavirus(PBV) | |
| Vibrio cholerae O1 | Salmonella | |
| Vibrio cholerae O139 | Shigella | |
| Vibrio cholerae CTX | Campylobacter laridis | |
| Vibrio cholerae ctxB | Campylobacter coli | |
| Vibrio parahaemolyticus | Yersinia pseudotuberculosis | |
| Vibrio parahaemolyticus TDH | Aeromonas hydrophila | |
| gene | Plesiomonas shigelloides | |
| Vibrio parahaemolyticus TRH | Vibrio mimicus | |
| gene | Vibrio vulnificus | |
| Vibrio parahaemolyticus TLH gene | Vibrio alginolyticus | |
| Vibrio parahaemolyticus ToxR | Vibrio fluvialis | |
| gene | Helicobacter pylori | |
| Staphylococcus aureus | Clostridium difficile | |
| SEA | Dysentery bacillus | |
| SEB | Proteus | |
| SEC | Enterobacter sakazakii | |

Infection related-Hepatitis virus

9

| | | | |
|------------|------------------------|------------|-----|
| HAV | HBV YMDD mutation | HCV | HEV |
| HBV | HBV mutations in the | HCV typing | |
| HBV typing | anterior C /BCP region | HDV | |

Infection related-Reproductive health

17

| | | | |
|---------------|--------|------------|----|
| HIV- | HPVB19 | UU | CT |
| I HIV- | RV | UU typing | TP |
| EB | TOX | HPV | |
| CMV | GBS | HPV typing | |
| HSV(I / II) | MH | NG | |

Infection related-Arthropod-borne disease

56

| | | | |
|--------------|------------------------|-------------------------|--------------------------|
| JEV | Lassa virus | Nam Dinh virus | Enterococcus faecium |
| DV universal | Brazilian haemorrhagic | Tibetan Circovirus | Enterobacter cloacae |
| DV I | fever virus | Akabane virus | Enterococcus faecalis |
| DV II | RVFV | Oya virus | Proteus mirabilis |
| DV III | NiV | GETV | Arenaviridae |
| DV IV | SLEV | Mangshi virus | Lassa virus |
| YFV | TBEV | Omono river virus | New Bunia virus |
| Zika virus | WEEV | Eridge virus | Variola Virus |
| CHIKV | EEEV | Menghai rhabdovirus | West Nile virus |
| SFTSV | VEEV | Guangping virus | Monkeypox Virus |
| WNV | RV | Crimean-Congo | Streptococcus agalactiae |
| XHFV | PRV | hemorrhagic fever virus | |
| HTNV | HTNV | CHIKV | |
| TOSV | Cardipino virus | RVFV | |
| MARV | Banna Virus | E.coli | |

Infection related-Parasite

14

| | | | |
|-----------------------|-----------------------------|-----------------------------|-----------------------|
| Plasmodium universal | Plasmodium malariae | Cryptosporidium | TOXO |
| Plasmodium falciparum | Plasmodium ovale (Curtisi) | Sparganum mansoni | Entamoeba histolytica |
| Plasmodium vivax | Plasmodium oval (Wallikeri) | Taenia solium | |
| Plasmodium ovale | Giardia lamblia | Angiostrongylus cantonensis | |

Tumor related-Copy number variation

2

HER2 copy number variation MET copy number variation

Tumor related-Methylation detection

12

| | | | |
|---------------------|---------------------|--------------------|-----------------------|
| Septin9 methylation | BMP6 methylation | SHOX2 methylation | DACH6 methylation |
| SDC2 methylation | P16 methylation | CPXM1 methylation | Cyclin D2 methylation |
| NDRG 4 methylation | RASSF1A methylation | HoxA10 methylation | PAX5 methylation |

| | | | |
|---------------------|--------------------|------------------|----------------|
| EGFR | - Exon2 Codon12,13 | -- Exon20 G1049G | - Exon18 D842V |
| - Exon19 del (LREA) | - Exon3 Codon59,61 | BRCA1 | - Exon12 |
| - Exon21 L858R | BRAF gene | - Exon11 | - Exon14 |
| - Exon18 G719X | - Exon15 V600E | - Exon2 | IDH1 |
| - Exon21 L861Q | - Exon15 V600D | - Exon20 | - R132C |
| - Exon20 T790M | - Exon15 V600K | - Exon5 | - R132G |
| - Exon20 ins | - Exon15 V600R | - Exon18 | - R132H |
| KRAS | MET | - Exon21 | JAK2 |
| - G12D | - Exon 14 | BRCA2 | - V617F |
| - G12V | HER2 | - Exon10 | MYD88 |
| - G12S | - Exon20 ins | - Exon11 | - L265P |
| - G12C | - Exon8 | C-KIT | TP53 |
| - G12A | - Exon17 V659E | - Exon11 | - R175H |
| - G12R | PIK3CA | - Exon9 | - R248L |
| - G13A | - Exon9 E542K | - Exon13 | - R273C |
| - G13D | - Exon9 E545Q | - Exon17 | - R273H |
| NRAS | - Exon20 H1047R | PDGFRA | |

Tumor related-Fusion gene detection

4

| | | | |
|------------|-------------|------------|----------------|
| ALK fusion | ROS1 fusion | RET fusion | BCR-ABL fusion |
|------------|-------------|------------|----------------|

Tumor related-Single gene disease

18

| | | | |
|----------------------|---------------------|----------------------|-----------------------|
| DMD | β thalassemia | Gal | GD |
| Hemophilia A | SMA | HLD | Mucopolysaccharidosis |
| Hemophilia B | CAH | PKD | type I |
| Hereditary deafness | PKU | Glycogenosis type II | Mucopolysaccharidosis |
| α thalassemia | Albino | Citrullinemia | type II |

Animal related-Animal disease

12

| | | | |
|-------|-------|------|---------------------|
| ASFV | PEV | PRV | PPR |
| CSFV | PCV-2 | PPV | NDV |
| PRRSV | FMDV | AHSV | Bovine tuberculosis |

Animal related-Zoonotic disease

10

| | | | |
|--------------------|------------------------|--------------------|----------------------|
| Yersinia pestis | Rickettsia prowazekii | Coxiella burnetii | Borrelia burgdorferi |
| Bacillus anthracis | Rickettsia mooseri | Bartonellahenselae | |
| Leptospira | Orientia tsutsugamushi | Brucella | |

Animal related-Pets

13

| | | | |
|------------------------|---------------------------|-------|-----|
| FPV | Chlamydia felis | CAV 2 | CCV |
| FHV | Cat tritrichomonas foetus | CPIV | |
| Dog and cat mycoplasma | TOX/Bartonella henselae | Bb | |
| FCV | CDV | CPV | |

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